

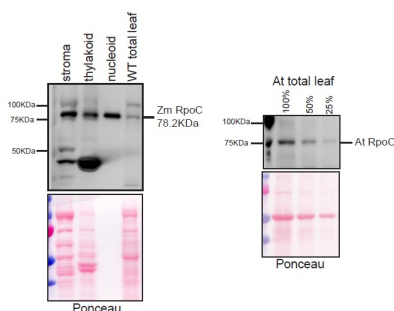
Product no **AS15 2868****Anti-RpoC1 | RNA polymerase beta' subunit (chloroplast)****Product information**

Immunogen	His-tagged, highly conserved fragment of <i>Zea mays</i> RpoC. UniProt: A0A059Q8F2
Host	Rabbit
Clonality	Polyclonal
Purity	Immunogen affinity purified serum in PBS pH 7.4.
Format	Lyophilized
Quantity	100 µg
Reconstitution	For reconstitution, add 50 µl of sterile water
Storage	Store lyophilized/reconstituted at -20°C; once reconstituted make aliquots to avoid repeated freeze-thaw cycles. Please remember to spin the tubes briefly prior to opening them to avoid any losses that might occur from material adhering to the cap or sides of the tube.

Application information**Recommended dilution** | 1 : 500-1 : 1000 (WB)**Expected | apparent MW** | 78 | 78 kDa**Confirmed reactivity** | *Arabidopsis thaliana*, *Zea mays*

Predicted reactivity | *Alloteropsis semialata*, *Aristida purpurea*, *Campanula americana*, *Chasmanthium latifolium*, *Chaetobromus involucratus* subsp. *dregeanus*, *Chionochloa macra*, *Coleataenia prionitis*, *Danthonia californica*, *Digitaria exilis*, *Echinochloa oryzicola*, *Eragrostis tef*, *Eremitis* sp., *Isachne distichophylla*, *Hilaria cenchroides*, *Hickelia madagascariensis*, *Microlaena stipoides*, *Miscanthus sinensis*, *Neololeba atra*, *Olmecca reflexa*, *Oryza sativa*, *Otatea acuminata*, *Panicum miliaceum*, *Pariana campestris*, *Phragmites australis*, *Potamophila parviflora*, *Puelia olyrififormis*, *Rhynchoryza subulata*, *Saccharum hybrid*, *Saccharum officinarum*, *Setaria viridis*, *Sorghum bicolor*, *Sorghum timorense*, *Sporobolus maritimus*, *Stipa purpurea*, *Triticum aestivum*, *Zoysia macrantha*

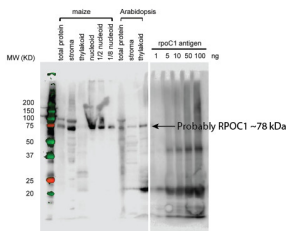
Species of your interest not listed? [Contact us](#)

Not reactive in | No confirmed exceptions from predicted reactivity are currently known**Additional information** | There may be occur a cross reacting band at 100 kDa in maize samples**Samples:**

Left panel: *Zea mays* extracts: whole leaf (5 ug), chloroplast nucleoid (0.1 ug), thylakoid (5 ug), or stroma (5 ug) as indicated MW markers are marked

5 µg/well of total protein extracted from *Arabidopsis thaliana* and *Zea mays* in 100 mM Tris-HCL pH7.5, 10% glycerol, 5mM EDTA and 2MM EGTA buffer and denatured in 66mM Tris-HCL pH6.8, 0.2% glycerol, 0.23% SDS, 600mM BME at 50°C/10 min. Samples were separated on 4-20 % SDS-PAGE Novex™ Tris-Glycine Mini Protein Gels (4–20%, 1.0 mm thick) and blotted overnight to nitrocellulose (0.22µm), using: wet transfer. Blot was blocked with 5 % milk for: 1h/RT with agitation. Blot was incubated in the primary antibody at a dilution of 1:500 for 2h/RT with agitation in 5% non-fat milk TBS-T with agitation. The antibody solution was decanted, and the blot was rinsed briefly 4 times in TBS-T for 5min each with agitation. Blot was incubated in matching secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:1000 in 5% non-fat milk TBS-T for 1h/RT with agitation. The blot was washed as above and developed with a following chemiluminescent detection reagent AgriseraECL SuperBright. Exposure time was 5 minutes.

Courtesy of Jie Shen and Dr. Alice Barkan, University of Oregon, USA



Stroma, thylakoid, nuclei and total protein from *Arabidopsis thaliana* and *Zea mays* were extracted with 50 mM HEPES-KOH pH 8, 330 mM sorbitol extraction buffer. Components were separated on 12% SDS-PAGE and blotted overnight to Nitrocellulose membrane using wet transfer. Blots were blocked with 5% non-fat milk for 1h at room temperature (RT) with agitation. Blot was incubated in the primary antibody at a dilution of 1: 500 for 2h at RT with agitation. The antibody solution was decanted and the blot was rinsed briefly twice, then washed once for 15 min and 3 times for 5 min in TBS-T at RT with agitation. Blot was incubated in secondary antibody (anti-rabbit IgG horse radish peroxidase conjugated) diluted to 1:10 000 in for 1h at RT with agitation. The blot was washed as above and developed for 5min with chemiluminescent detection reagent of extreme low femtomogram range. Exposure time was 120 seconds.

Courtesy of Jie Shen and Dr. Alice Barkan, University of Oregon, USA